Animal cells bioassays and bioproducts

Introduction

Biological assays are methods for the estimation of nature, constitution, or potency of a material by means of the reaction that follows its application to living matter. Bioassay is defined as estimation or determination of concentration or potency of physical, chemical or biological agents by means of measuring and comparing the magnitude of the response of the test with that of standard over a suitable biological system under standard set of conditions [1,2]. An assay is a form of biological experiment; but the interest lies in comparing the potencies of treatments on an agreed scale, instead of in comparing the magnitude of effects of different treatments. Biological assays or biological standardizations or simply bioassays are methods used for estimation of the potency of substances by observing their pharmacological effects on living animals (in vivo) or isolated tissues (in vitro) and comparing the effect of these substances of unknown potency to the effect of a standard.

Bioassays are based upon the use of biological responses as detection system for biologically active substances. In the simplest form it is used to assay the presence (and concentration) of a particular substance by comparison with a known amount of the same substance. Both are procedures by which the potency or the nature of a substance is estimated by studying its effects on living matter. Bioassay is a procedure for the determination of the concentration of a particular constitution of a mixture [5-8].

Structure of biological assays

The typical bioassay involves a stimulus applied to a subject. Application of stimulus is followed by a change in some measurable characteristic of the subject, the magnitude of the change being dependent upon the dose. The intensity of the stimulus is varied by

using the various Doses by the analyst.

Principle of Bioassay

Active principle to be assayed should show the same measured response in all animal species [9]. Bioassay involves the comparison of the main pharmacological response of the unknown preparation with that of the standard [10-14]. The method selected should be reliable, sensitive, and reproducible and should minimize errors due to biological variation and methodology. The degree of pharmacological response produced should be reproducible under identical conditions. The reference standard and test sample should have same pharmacological effect and mode of action, so that their DRC curve run parallel and their potency ratio can be calculated [15-18]. Activity assayed should be the activity of interest; Individual variations must be minimized/accounted for [19]. Bioassay might measure a diff aspect of the same substance compared to chemical assay.

Types of Bioassays

There are three main types of bioassays (other than qualitative assays) [27]

- 1. Direct Assays
- 2. Indirect Assays based upon quantitative responses
- 3. Indirect Assays based upon quantal responses ("all or none")

Direct Assay

Doses of the standard and test preparations are sufficient to produce a specified response, and can be directly measured.

Indirect Assay

In indirect bio-assays the relationship between the dose and response of each preparation is first ascertained. Then the dose corresponding to a given response is obtained from the relation for each preparation separately [29].

Quantal Assay

This response is in the form of "all or none" means no response or maximum response. These can be biossayed by end point method. Predetermined response is measured which is produced by threshold effect. Quantal Responses are population response based on an all-or-nothing (0 or 1 – presence or absence) response such as death [30-34].

Concentration of Unknown = Dose of the Standard Dose of the Test×Concentration of Standard

Graded Assay

It is proportional to the dose and response may lie between no response and maximum response [28]. Graded Responses can be any type of measured responses in isolated tissues in particular, but also in whole animals. Such responses are infinitely graded and there are a large number of them. Examples include contractions of muscle, blood pressure, blood sugar concentrations, etc. [35]

Matching Method

In this type of assay the test substance and the standard are applied and the responses obtained are matched by a trial and error process until they produce equal effects [36-38]. This may also limit to analytical dilution assay, as the assay involves the determination of the factor by which the test substance is diluted or concentrated in order to produce response that is equal to that of known amount of the standard drug [39-40]. Its advantage is that it does not depend on the assumption of a dose-response relationship. The main disadvantages are that it is purely subjective, and experimental errors cannot be determined from the assay. It gives no indication or the parallelism of the dose-response curves of the standard drug and test substance, and hence the qualitative differences, as the effects are matched at only one dose level. [41-46] **Advantages:**

Quick and easy; useful when one is has many samples to test and a semiqualitative

answer is sufficient. Disadvantages: Inherently lacks precision, no accuracy, no D-R data – particularly no data regarding slope. The data is not easily statistically analyzed and probably should not be so analyzed [47].

Bracketing Method

Bracketing bioassay is performed by selecting two standard doses, which will give a close bracket on either side of the response produced by the unknown. The working dose of standard is first determined in the sensitive part of dose-response curve, that is, a dose that will approximately produce 50% of the maximal concentration. The dose of the standard drug is kept constant throughout the experiment, in order to have some idea about the change in the sensitivity of tissue with time. [48-53] The standard drug is added at fixed intervals but alternating with the test so that each response produced by a dose of test substance is bracketed by responses produced by the dose of standard. The response of test substance is bracketed between two responses of the standard. Close bracketing gives more accurate results. [54-56]

Interpolation Method

This is a simplest form of graded response assay and involves no statistical data and many calculations. In this assay the dose response curve is fist obtained from different doses of standard ach solution. The concentration of unknown is then read from the standard graph. [57-62] Interpolation method of bioassay is less time consuming and yet reliable compare to matching type of bioassay. One of the main advantages of this essay is that the sensitivity of the tissue is first determined by prior plotting of a dose response curve with a known agonist as in the case with acetylcholine. If the linearity of curve is good, one can do very accurate estimate of the test substance unknown sample.

Characteristics

A good bioassay should has the following characteristics

Sensitivity- ability to detect smallest concentration

Specificity-the response which is being measured should be specific

Reproducibility-same observations by using different instruments and operators, over longer period of time

Stability-sensitivity of preparation should be constant and stable

Availability-the particular tissue or cell should be easily available

Advantages

There are a number of advantages to including Maximum Tolerated Dose(MTD) in longterm animals bioassays

Interspecies comparison

When bioassays are conducted in more than one animal specie, use of MTD provides a consistent basis for interspecies comparisons

Sensitivity

The greater sensitivity of MTD are more likely to give positive or negative results than can be starting point for structural activity correlation analysis.

Preparations

Several preparations can be obtained from single animal cell

Cheap

Animal cells bioassays is a very cheap and less time consuming method than other methods

Disadvantages

Where there are advantages of animal cells bioassays ,there are some disadvantages of bioassays too

Non-specificity

Bioassay is non-specific and provides no information about biochemical and physiological mechanism during tumorproduction

Toxicity

It induces toxicity that leads to change in food consumptions, cytotoxicity in specific organs and hormonal imbalance.

Cross contamination

There is high possibility of cross-contamination of different types of cells while working with bioassays

Identification

Identification of cell type is often difficult in most of the cases, the markers proteins are not expressed under in-vitro conditions

Applications

The applications of animal cells bioassays are as follows

Detection and isolation of proteins

Bioassays is used for the detection and isolation of proteins such as, somatotrophin, insulin-like growth factors (somatomedins), insulin and transferrin etc

Detection of muscle growth factors

Radioimmunoassays cannot be used effectively for the detection and characterization of unknown and poorly characterized muscle growth factor.so for such detection bioassays are capable and reliable method that detects those factors that influencing the muscle growth.

Potency of agents

Animal cells bioassay is used to estimate the potency of agents and their effects.

Cytotoxicity studies

Bioassay is used to check the in-vitro toxicity of compounds or drugs in animals cells.

Therapeutic products

Bioassays are used to establish the acticity of therapeutic products.

REFERENCES

1. Agatonovic-Kustrin S et al. Thin-Layer Chromatography - Bioassay as Powerful Tool for Rapid Identification of Bioactive Components in Botanical Extracts. Mod Chem appl. 2015; 3:e120.

10. Singh SV and Swami VK. Utilization of Distillery Industry Wastewater as Liquid Biofertilizer: Seed Bioassay Test for Feasibility and Toxicity Assessment. IJIRSET. 2014; 3: 17021-17027.

11. Lourenço FR. Simple Estimation of Uncertainty in the Quantification of Cefazolin by HPLC and Bioassay. J Chromat Separation Techniq. 2012; 3:153.

12. de Baptista Neto Õet al. An Alternative Methodology for Determination of Cephamycin Cfrom Fermentation Broth. J Chromat Separation Techniq. 2012; 3:130.

13. Lewis DR and Liu DJ. Direct Measurement of Lipase Inhibition by Orlistat Using a Dissolution Linked In Vitro Assay. Clinic Pharmacol Biopharm. 2012; 1:103.

14. Corrêa JCR et al. AStability Study of Fluconazole Applying Validated Bioassay and Stability-Indicating LC Methods. J Anal Bioanal Tech. 2011; 2:126.

15. Hiwasa Tetal. Functional Similarity of Anticancer Drugs by MTT Bioassay. J Cancer Sci Ther. 2011; 3:250-255.

16. Zakeri-Milani P et al. Pharmacokinetic Study of Two Macrolide Antibiotic Oral Suspensions Using an Optimized Bioassay Procedure. J Bioequiv Availab. 2010; 2: 111-

115.

17. Brayden DJ. Intestinal epithelial permeation enhancers: High content screening reveals sub-lethal cellular effects of sodium caprate. Pharmaceut Anal Acta 2013; 4:2

18. Gupta A et al. A Randomised Double-Blinded Dose Response Study of the Fentanyl with Hyperbaric Ropivacaine in Cesarean Section. J Anesth Clin Res. 2014; 5:467.

19. Turaka A. Dose Response to Radiation Therapy for Primary Ocular Lymphomas. OMICS J Radiology. 2013;2:e118.

2. Hemanta MR et al. Analysis of Traditional Food Additive Kolakhar for its Physico-Chemical Parameters and Antimicrobial Activity. J Food Process Technol. 2014; 5:387.

20. Bhandare N and Mendenhall WM. A Literature Review of Late Complications of Radiation Therapy for Head and Neck Cancers: Incidence and Dose Response. J Nucl Med Radiat Ther. 2012; S2:009.

21. Shintani H. Methods of Rapid Microbiological Assay and Their Application to Pharmaceutical and Medical Device Fabrication. Biochem Physiol. 2014; 3:141.

22. Matías J et al. Preliminary Studies on a Derivative Verotoxin as Oral Adjuvant. J Vaccines Vaccin. 2015; 6:279.

23. Madar-Balakirski N et al. Measurement of Cellular Immunity to Influenza Vaccination in Rheumatoid Arthritis; Comparison of Three Assays. J Vaccines Vaccin. 2015; 6: 278.

24. Zheng T et al. Adjuvanticity of a Synthetic Phosphatidylinositol Dimannoside to a Subvirion Influenza Vaccine in an Influenza Mouse Model. J Vaccines Vaccin. 2015; 6: 277.

25. Huang J et al. Biochemical and Immunological Characterizations of the Receptor Binding Domain of C. difficile Toxin B. J Vaccines Vaccin. 2015; 6: 276.

26. Gimenez-Sanchez F et al. A Matched CaseControl Study Measuring the Effectiveness of the Rotavirus Vaccines to Prevent Gastroenteritis Hospitalizations. J Vaccines Vaccin. 2015; 6:275.

27. Qi F et al. Immune-Based Modulation of Adult Hippocampal Neurogenesis, Link to Systemic Th1/Th2 Balance. J Vaccines Vaccin. 2015; 6: 274.

28. Lo Giudice D et al. Human Papillomavirus Vaccination Coverage among Adolescents Living in Southern Italy. J Vaccines Vaccin. 2015; 6: 273.

29. Mshelbwala PP et al. Evaluation of Two Rapid Diagnostic Tests for Rabies Diagnosis under Field and Laboratory Conditions in Nigeria. J Vaccines Vaccin. 2015; 6: 272.

3. Priac A et al. Ecotoxicity Evaluation of Industrial Discharge Waters and Metallic Solutions using Two Organisms (Lactuca sativa and Daphnia magna). J Pollut Eff Cont. 2014; 2:117

30. Alasil SM and Kutty PK. Breastfeeding as a Tool that Empowers Infant Immunity through Maternal Vaccination. J Vaccines Vaccin. 2015; 6: 271.

31. Tsuda B, Kametani Y et al. The Effect of Peptide Treatment on the HLA-Binding and Antibody Production in Peripheral Blood Mononuclear Cells Obtained from Japanese Breast Cancer Patients. J Vaccines Vaccin. 2015; 6: 270.

32. Selvaraj J et al. Analysis of Alternative Purification of Beta-Propiolactone Inactivated, Tangential Flow Filtration Concentrated Vero Cell Derived Rabies Vaccine. J Vaccines Vaccin. 2015; 6:269.

33. Berera D and Thompson KM. Medical Student Knowledge, Attitudes, and Practices Regarding Immunization. J Vaccines Vaccin. 2015; 6: 268.

34. Nicolellis F. An Early Report on a Local Project about Primary Health Care to Improve the Communication and the Compliance in the Elderly for Vaccine Campaigns against Influenza. J Vaccines Vaccin. 2015; 6: 267.

35. Calderon-Gonzalez G et al. A Dendritic Cell-Targeted Vaccine Loaded with Glyceraldehyde-3- Phosphate Dehydrogenase Peptides Proposed for Individuals at High Risk of Listeriosis . J Vaccines Vaccin. 2015; 6: 266.

36. Cirelli E et al. Retinoic Acid Promotes Mucosal and Systemic Immune Responses after Mucosal Priming and Systemic Boosting in Mice. J Vaccines Vaccin. 2015; 6: 265.

37. Chaves Sp et al. Serine Proteases and Vaccines against Leishmaniasis: A Dual Role . J Vaccines Vaccin. 2015; 6:264.

38. Otolorin GR et al. A Review on Human Deaths Associated with Rabies in Nigeria. J Vaccines Vaccin. 2015; 6:262.

39. Megid J. Vaccinia Virus: It's Use in Smallpox Vaccine and Epidemiology. J Vaccines Vaccin. 2015; 6:261.

4. Hu B et al. Bioassay-guided Isolation of the Antidiabetic Active Principle from Salvia miltiorrhiza and its Stimulatory Effects on Glucose Uptake Using 3T3-L1 Adipocytes. Med chem. 2014; 4:592-597.

40. Wertheimer A. Therapeutic Vaccines: A New Revolution. J Pharmacovigi. 2015, 3:1

41. Nerome K et al. The Usefulness of an Influenza Virus-Like Particle (VLP) Vaccine Produced in Silkworm Pupae and Virosomes and Liposomes Prepared by Chemical Means: From Virosome to VLP and the Future of Vaccines. J Gastrointest Dig Syst. 2015; 5:256.

42. McCullough KC et al. Dendritic Cell Targets for Self-Replicating RNA Vaccines. J Blood Lymph. 2015; 5: 132.

43. Fernandes FF et al. Molecular Modeling of Heat Shock Protein of 60-Kda from Paracoccidioides Brasiliensis: The First in silico Structural Model of a Fungal Hsp60. J Comput Sci Syst Biol. 2014; 8: 241-244.

44. Tomisaka M et al. Overcoming the Japanese "Vaccine Gap": An Analysis of Medical Leaders" Witness. J Vaccines Vaccin. 2015; 6: 263.

45. Akbayram et al. Vaccination Associated Acute Immune Thrombocytopenic Purpurai Children. J Vaccines Vaccin. 2014; 5:6.

46. Bhardwaj V et al. AgNPs-Based Label-Free Colloidal SERS Nanosensor for the Rapid

and Sensitive Detection of Stress-Proteins Expressed in Response to Environmental-Toxins. J Biosens Bioelectron. 2013; S12: 005.

47. Bhosle P and Vaibhav S. Conotoxins: Possible Therapeutic Measure for Huntingtons Disease. J Neurol Disord. 2013; 1: 129.

48. Vasconcelos V. Emergent Marine Toxins in Europe: is there a new Invasion? J Marine Sci Res Dev. 2013; 3: e117.

49. Fernández H. Mycotoxins Quantification in the Food System: Is there Any Contribution from Electrochemical Biosensors? J Biosens Bioelectron. 2013; 4: e121.

5. Zainal B et al. Anticancer Agents from Non-Edible Parts of Theobroma cacao. Nat Prod Chem Res. 2014; 2:134.

50. Martin K and Nashar TO. E. coli Heat-labile Enterotoxin B Subunit as a Platform for the Delivery of HIV Gag p24 Antigen. J Clin Cell Immunol. 2013; 4: 140.

51. Zagatto PA and Ferrão-Filho AdS. Acute Effects of a Cylindrospermopsis Raciborskii (Cyanobacteria) Strain on Mouse, Daphnia and Fish. J Ecosyst Ecogr. 2013; 3: 121.

52. Sasani F et al. The Relationship between Microscopic Lesions and Different Types of Clostridium perfringens and their Related Toxins by Sandwich ELISA in Cattle. J Microb Biochem Technol. 2013; 5:034-038.

53. Kumar Y and Kurcheti PP. Effect of Liver Biotoxins of Certain Marine Fishes on Mouse Cell Culture. J Marine Sci Res Dev. 2013; 3: 117.

54. Shi H et al. Assessment and Removal of Emerging Water Contaminants. J Environ Anal Toxicol. 2012; S2:003.

55. Anderson PD and Bokor G. Conotoxins: Potential Weapons from the Sea. J Bioterr Biodef. 2012; 3:120

56. Shintani H. Inactivation of Prion and Entotoxins by Nitrogen Gas Plasma Exposure. Pharmaceut Anal Acta. 2012; 3:177.

57. Mokhtar G and Naoyuki F. Microfiltration, Nano-filtration and Reverse Osmosis for the Removal of Toxins (LPS Endotoxins) from Wastewater. J Memb Sci Technol. 2012; 2:118.

58. Masters RD. Acetylcholines, Toxins and Human Behavior. J Clinic Toxicol. 2012; S6:004.

59. Singh MK et al. Applications of Rat Brain Synaptic Vesicle Proteins for Sensitive and Specific Detection of Botulinum Neurotoxins. J Bioterr Biodef. 2012; S2:008.

6. Ado K et al. Fundulopanchax gardneri Test: A Convenient Method of Bioassay for Active Constituents of Natural Products. Nat Prod Chem Res. 2014; 2:133.

60. Gong X et al. Determination of 15 Mycotoxins in Foods and Feeds Using High Performance Liquid Chromatography-Tandem Mass Spectrometry with Gel Permeation Chromatography Combined QuEChERS Purification. J Chromat Separation Techniq. 2012; 3:125.